


Schedule of Accreditation

issued by

United Kingdom Accreditation Service

21 - 47 High Street, Feltham, Middlesex, TW13 4UN, UK

 2293 Accredited to ISO/IEC 17025:2005	Cefas, Food Safety	
	Issue No: 026 Issue date: 23 April 2012	
	The Cefas Weymouth Laboratory The Nothe Barrack Road Weymouth DT4 8UB	Contact: Dr D Lees Tel: +44 (0)1305 206600 Fax: +44 (0)1305 206601 E-Mail: D.J.Page@cefass.co.uk Website: www.cefass.co.uk
Testing performed by the Organisation at the locations specified below		

Locations covered by the organisation and their relevant activities

Laboratory locations:

Location details	Activity	Location code
Address CEFAS Weymouth Laboratory Barrack Road The Nothe Weymouth Dorset DT4 8UB Local contact Ms D Page Tel: +44 (0)1305 206600 Fax: +44 (0)1305 206601 Email: D.J.Page@cefass.co.uk Website: www.cefass.co.uk	All	A

Site activities performed away from the locations listed above:

Location details	Activity	Location code
Fish Farms	Sampling of fish tissues for subsequent virological and bacterial testing	B



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DETAIL OF ACCREDITATION

Materials/Products tested	Type of test/Properties measured/Range of measurement	Standard specifications/ Equipment/Techniques used	Location Code
FOOD AND FOOD PRODUCTS - BIVALVE MOLLUSCS Pacific oysters, native oysters, cockles, hard clams, whole king scallops, mussels Cockles, razor & hard clams, mussels, pacific oysters, native oysters, king and queen scallops, razor, hard, surf, manila, otter and carpet clams Mussels, pacific oysters, native oysters, common cockles, razor clams, king scallops and queen scallops	<u>Chemical Analysis</u> Detection of Domoic acid (amnesic shellfish poisoning toxin, ASP),	Documented in-house methods: SOPs 1180, 1744, 1200, 1243 and 1723, by HPLC-UV	A
	Paralytic shellfish poisoning toxin (PSP): Qualitative test	SOPs 1180, 1181, 1743, 1744, 1745 and 1746, by HPLC-FLD	A
	Paralytic shellfish poisoning toxin (PSP): Qualitative pre-screen and quantitative test	SOPs 1180, 1744, 1784, 1785, 1788 by HPLC-FLD based on AOAC Official method 2005.06	A
	Lipophilic Marine Toxins (DSP) : Quantitative test	SOPs 2098,2097 and 2096 by LC-MS/MS	A
Bivalve mollusc digestive gland	<u>Molecular Biology</u> Detection of norovirus genogroup I and II RNA	SOP 1330 by real-time reverse transcriptase PCR using Mx3005P Stratagene thermocycler	A
FOOD AND FOOD PRODUCTS - BIVALVE MOLLUSCS	<u>Microbiological Tests</u> Enumeration: <i>Escherichia coli</i>	SOP 1175, based on ISO/TS 16649-3:2005	A
	Detection: <i>Salmonella</i> spp (excluding <i>S. typhi</i>)	SOP 1176	A
	<i>Vibrio parahaemolyticus</i>	SOP 1333	A



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<p>FOOD AND FOOD PRODUCTS (cont'd)</p> <p>- BIVALVE MOLLUSCS (cont'd)</p> <p>HPLC pre-screened common mussels, pacific oysters, native oysters, common cockles and whole king scallops. Unscreened bivalve molluscs of species other than the above</p> <p>FISH, Bivalve Mollusc and Crustacean tissues</p> <p>Fish</p> <p>Bivalve Mollusc</p>	<p><u>Microbiological Tests</u> (cont'd)</p> <p><u>Bioassays</u></p> <p>Paralytic shellfish poisoning toxin (PSP), quantitative test</p> <p><u>Sampling</u></p> <p>For tissues submitted for virological, molecular, bacterial and histopathological diagnostic testing to undertake a full disease screen, including screening for all diseases affecting fish listed in Council Directive 2006/88/EC and the Aquatic Animal Health (England and Wales) Regulations 2009</p> <p>For tissues submitted for histopathological diagnostic testing to undertake a full disease screen, including screening for all diseases affecting bivalve molluscs listed in Council Directive 2006/88/EC and the Aquatic Animal Health (England and Wales) Regulations 2009</p>	<p>SOPs 1180, 1181 and 1183</p> <p>Documented in-house methods in accordance with the following:</p> <p>SOP 2020 - Fish sampling methodology in accordance with: Council Directive 2006/88/EC The Aquatic Animal Health (England and Wales) Regulations 2009</p> <p>SOP 2021 - Mollusc Sampling Methodology</p> <p>In accordance with: Council Directive 2006/88/EC The Aquatic Animal Health (England and Wales) Regulations 2009</p>	<p>A</p> <p>A,B</p> <p>A,B</p>



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FISH, Bivalve Mollusc and Crustacean tissues (cont'd)	<u>Sampling</u> (sampling)	Documented in-house methods in accordance with the following:	
Crustaceans	For tissues and fluids submitted for molecular, bacterial and histopathological diagnostic testing to undertake a full disease screen, including screening for all diseases affecting crustaceans listed in Council Directive 2006/88/EC and the Aquatic Animal Health (England and Wales) Regulations 2009	SOP 2022 - Crustacea Sampling Methodology In accordance with: Council Directive 2006/88/EC The Aquatic Animal Health (England and Wales) Regulations 2009	A,B
	<u>Testing</u>	Documented in-house methods in accordance with the following:	
	<u>Virological Tests</u>		
Brain, Kidney and Spleen	Isolation and identification of Viral Haemorrhagic Septicaemia (VHS) virus	SOP's 1248, 1279, 1125 and 1259 by tissue culture and ELISA in accordance with Commission Decision 2001/183/EC, Council Directive 2006/88/EC (based on OIE Manual of Diagnostic Tests for Aquatic Animals, 2009)	A
	Isolation and identification of Infectious Haematopoietic Necrosis (IHN) virus	SOP's 1248, 1279, 1125 and 1263 by tissue culture and IFAT in accordance with Commission Decision 2001/183/EC, Council Directive 2006/88/EC (based on OIE Manual of Diagnostic Tests for Aquatic Animals, 2009)	A
	Isolation and identification of spring Viraemia of Carp (SVC) virus	SOP's 1248, 1279, 1125 and 1259 by tissue culture and ELISA in accordance with Commission Decision 2001/183/EC, Council Directive 2006/88/EC (based on OIE Manual of Diagnostic Tests for Aquatic Animals, 2009)	A



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FISH, Bivalve Mollusc and Crustacean tissues (cont'd)	<u>Testing</u> (cont'd) <u>Virological Tests</u> (cont'd) Virus isolation procedures for EHNV- epizootic haematopoietic necrosis virus: Inoculation of samples onto tissue culture cells (BF2 and EPC cells) in fish tissue samples (homogenate supernatants)	Documented in-house methods in accordance with the following: SOP's 1248, 1279 and 1125 in accordance with Council Directive 2006/88/EC (based on OIE Manual of Diagnostic Tests for Aquatic Animals, 2009)	A
Fish, shellfish, crustacean tissues	Detection and Identification of Spring Viraemia of Carp virus (SVCV) RNA	SOPs 2063,1245, 2008, 2006,2007, 1537, 1742 and 1546 by RT-PCR. Guidance from the Aquatic Animal Health Directive - Council Directive 2006/88 (based on OIE Manual of Diagnostic tests for Aquatic Animals 2006)	A
Fish, shellfish, crustacean tissues	Detection and Identification of Koi Herpes virus (KHV) DNA	SOPs 1493,1245, 2008, 2006,2007, 1742 and 1546 by PCR. Guidance from the Aquatic Animal Health Directive - Council Directive 2006/88 (based on OIE Manual of Diagnostic tests for Aquatic Animals 2006)	A
Fish tissue and homogenates	Virus isolation procedures for ISAV – infectious salmon anaemia virus: inoculation of samples onto tissue culture cells (TO and SHK cells) and confirmation by IFAT <u>Histopathological Tests</u>	SOPs 1248, 12798, 1125,1263 & 1268 using inverted epifluorescent microscopy and IFAT based on OIE Manual of Diagnostic Tests for Aquatic Animals 2009	A
Bivalve molluscs	Screening for parasites	SOP 1681 in accordance with Commission Decision 2002/878/EC by haematoxylin and eosin staining	A
Crustacean Tissues	Detection of pathogens and notifiable diseases	SOP 1795 by Haematoxylin and Eosin staining using light microscopy	A



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FISH, Bivalve Mollusc and Crustacean tissues (cont'd)	<u>Testing (cont'd)</u>		
Salmonids fins	<u>Parasitology Tests</u> Identification of Gyrodactylus salaris	Documented in-house methods in accordance with the following: SOP 1735 by light microscopy	A
Fish, Shellfish and Crustacean tissue	<u>Pathology Tests</u> Detection and identification of viruses, bacteria, fungi and parasites	Documented in-house methods in accordance with the following: SOPs 1245, 2008, 2006,2007, 1537, 1742,1546 by robotic extraction, conventional PCR including RT-PCR and sequence analysis using ABI 3130 analyser under Flexible Scope SOP 2016	A
END			